



Phytochemical and pharmacological potential of *Artemisia absinthium* Linn. and *Artemisia asiatica* Nakai : A Review

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ABSTRACT

A world health organization survey indicated that about 75-80% of the world's populations rely on non-conventional medicine, mainly of herbal sources, in their primary healthcare. There has been an explosion of scientific information concerning plants, crude plant extracts and various substances from plants as medicinal agents during last 30-40 years. Although herbal medicine has existed since the dawn of time, our knowledge of how plants actually affect human physiology remains largely unexplored. Numbers of plants are claiming various medicinal uses and many researches are going on in this view. They are believed to be much safer and proved elixir in the treatment of various ailments. The genus *Artemisia* consists of about 500 species, occurring throughout the world. Among the different species of *Artemisia*, *A. absinthium* and *A. asiatica* have a vast range of biological activities including cytotoxic, antihepatotoxic, antibacterial, antifungal, antioxidant, antimalarial etc. Terpenoids, flavonoids, coumarins, caffeoylquinic acids, sterols and acetylenes constitute major classes of phytoconstituents of the genus *Artemisia*. The present review comprises the phytochemical, ethnopharmacological and pharmacological reports of *A. absinthium* and *A. asiatica*. The future scopes of these plants have been emphasized with a view to isolate bioactive moieties which could be used for multifarious biological activities.

Keywords: *Artemisia absinthium*, *Artemisia asiatica*, Antioxidant, Antimicrobial, Terpenoids

INTRODUCTION

Plants play a vital role in maintaining human health and contribute towards improvement of human life. They are important components of medicines, cosmetics, dyes, beverages etc. Plants have been one of the important sources of medicines even since the dawn of human civilization. In spite of tremendous development in the field of allopathy during the 20th century, plants still remain one of the major sources of drug in the modern as well as traditional system of medicine throughout the world. Over 60% of all pharmaceuticals are plant-based^[1]. Plants are considered as state-of-art chemical laboratories capable of biosynthesizing number of biomolecules of different chemical classes. Many of these are proved to be precursors for development of other drugs^[2]. India is one of the most medicoculturally diverse countries in the world where the medicinal plant sector is part of a time-honored tradition that is respected even today. Ethno-botanical and ethno-pharmacological studies on such plants continue to attract investigators throughout the world. In current scenario, focus on plant research has increased all over the world and a large body of evidence has collected to show immense potential of

medicinal plants used in various traditional systems^[3]. The present review emphasizes the traditional uses, phytochemistry and therapeutic potential of *A. absinthium* and *A. asiatica*. Through this review, authors hope to attract the attention of natural product researchers throughout the world to focus on the unexplored potential of *Artemisia* species.

The Genus *Artemisia*

The genus *Artemisia* belongs to a useful group of aromatic and medicinal plants. It is one of the largest and most widely distributed genera of the family Astraceae. It is a heterogenous genus, consisting over 450 diverse species distributed mainly in the temperate zone of Asia, Europe and North America. These species are perennial, biennial and annual herbs or small shrubs^[4-7].

Ethnopharmacology

A. absinthium has been traditionally used as an anthelmintic, antiseptic, antispasmodic, febrifuge, stomachic, cardiac stimulant, for the restoration of declining mental function and inflammation of the liver, and to improve memory^[8-11]. Traditional Chinese medicine practitioners use the plant for treating acute bacillary dysentery, cancers and neurodegenerative diseases^[12-15].

Extracts of the whole herb of *A. asiatica* have been used in

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traditional oriental medicine for the treatment of inflammation, cancer, infections and ulcerogenic diseases [16-19].

Phytochemistry

Exhaustive literature survey on phytochemical reports of *A. absinthium* and *A. asiatica* reveals that they comprise mainly terpenoids, flavonoids, coumarins, polyphenolics, caffeoylquinic acids, sterols and acetylenes. Both species are rich in terpenoids. Table-1 summarizes the phytoconstituents of *A. absinthium* and *A. asiatica*.

Pharmacological Reports

A. absinthium

Khattak et al. [43] reported that hexane-, chloroform-, and water-soluble extracts of *A. absinthium* exhibit antipyretic activity against subcutaneous yeast injections in rabbits. No toxic effects were documented for the plant extract at doses up to 1.6 g/kg. The essential oils distilled from the aerial parts of *A. absinthium* inhibited *in vitro* growth of *Candida albicans* and *Saccharomyces cerevisiae* var. *chevalieri* [22]. 5,6,3',5'-tetramethoxy 7,4'-hydroxyflavone, a flavone isolated from *A. absinthium* has been reported to exhibit *in vitro* anti-inflammatory activity as evidenced by inhibition of cyclo-oxygenase-2 (COX-2), prostaglandin (E-2 and PGE-2) and nitric oxide in lipopolysaccharide-stimulated RAW 264.7 cells [31]. *A. absinthium* has been studied for cognitive enhancement because of its nicotinic and muscarinic receptor activity (IC 50 concentration of less than 1 mg/mL) in homogenates of human cerebral cortical membranes [44]. The intoxicating effects of thujone were believed to activate receptors responsible for marijuana intoxication; however, thujone exhibited low affinity for rat cannabinoid receptors [45]. Methanol extract of *A. absinthium* enhanced neurite outgrowth induced by nerve growth factor and PC12D cells [46]. Free-radical scavenging activity of *A. absinthium* extracts has been reported [47]. The antioxidative activity was tested by measuring their ability to scavenge stable 2,2-diphenyl-1-picrylhydrazyl free radical and reactive hydroxyl radical during the Fenton reaction trapped by 5,5-dimethyl-1-pyrroline-N-oxide, using electron spin resonance spectroscopy. Aqueous methanolic extract of *A. absinthium* exhibited hepatoprotective effect against acetaminophen- and carbon tetrachloride-induced hepatic damage [12]. Recently it has been reported that crude aqueous extract and crude ethanolic extract of the aerial parts of *A. absinthium* exhibit anthelm-

intic activity in comparison to albendazole against the gastrointestinal nematodes of sheep [48]. Valdes et al. [49] reported that ethanol extract of *A. absinthium* exhibit antiprotozoal potential against *Trypanosoma brucei*, *Trypanosoma cruzi*, *Leishmania infantum* and *Plasmodium falciparum*; and antifungal activity against *Microsporium canis* and *Candida albicans*. Methanol extract of the aerial parts of *A. absinthium* at a dose of 300 mg/kg found effective against a trichinellosis (*Trichinella spiralis*) in rats [50]. Artemisetin isolated from *A. absinthium* exhibited marked antitumor activity against melanoma B16, but only weakly retarded growth of *Pliss lymphosarcoma* [51]. Chronic use of *A. absinthium* has been reported to have some neurotoxic effect due to presence of thujone and its derivatives [52].

A. asiatica

Ryu et al. [53] reported that DA-9601, a standardized extract of *A. asiatica* (10, 30, or 100 mg/kg, intragastrically) exhibit hepatoprotective effect on liver damage induced by acetaminophen and carbon tetrachloride. In another study, DA-9601 (10% for 15 week) exhibited chemopreventive effects against azoxymethane-initiated and dextran sulfate sodium-promoted mouse colon carcinogenesis. In addition, DA-9601 treatment also suppressed the expression of COX-2 and inducible nitric oxide synthetase as well as nuclear factor (NF)-kappa-B DNA binding in the colonic tissues [18]. Moreover, DA-9601 also reported to suppress the airway allergic inflammation via regulation of various cellular molecules expressed by MAP kinases/NF-Kappa-B pathway [54]. Artemisolide, as NF-kappa-B inhibitor, isolated from *A. asiatica* inhibited NF-kappa-B transcriptional activity in lipopolysaccharide-stimulated macrophages RAW 264.7 (IC 50 value 5.8 µM) [17]. DA-9601 has been reported to inhibit increased susceptibility of ethanol-treated gastric mucosa to naproxen [55].

Eupatilin (5,7-dihydroxy-3,4,6-trimethoxyflavone), a pharmacologically active ingredient isolated from *A. asiatica*, has been evaluated for apoptosis-inducing capability in cultured human promyelocytic leukemia (HL-60) cells. Eupatilin exhibited concentration-dependent inhibitory effects on viability and DNA synthesis capability of HL-60 cells [16]. Park et al. [56] reported that pre-treatment of *A. asiatica* extract (30 or 100 mg/kg), significantly attenuate gastric injury against alcohol-induced damage in gastric mucosa of rats, by significantly decreasing lipid peroxidation, preventing glutathione depletion, and inhibiting cytochrome 2E1 ethanol-metabolizing enzyme.

Table 1: Phytoconstituents of *A. absinthium* and *A. asiatica*.

Species	Phytoconstituents
<i>A. absinthium</i>	Essential oil containing chamazulene, nuciferol butanoate, nuciferol propionate, caryophyllene oxide, phellandrene, pinene, azulene [20], (Z)-thujone, (E)-thujone [5,11], myrcene, trans-sabinyl acetate [21], cis- and trans-epoxyocymenes, chrysanthenyl acetate [20,22], thujyl alcohol, nerol, isothujyl acetate [23], prochamazulenogen [24], β-pinene, hydrocarbon monoterpenes [25], sabinene, 1,8-cineole, <i>Artemisia</i> ketone, linalool, trans-verbenol, carvone, curcumene, neryl butyrate, neryl 2-methylbutanoate, neryl 3-methylbutanoate [26], sesquiterpene lactones arabsin, artabin, ketopelenolide, santonin related lactones [27]; tannins [28]; carotenoids ; lignans [29]; glucosides absinthin, anabsinthin [30]; phenolic compounds [5], flavonoid 5,6,3',5'-tetramethoxy 7,4'-hydroxyflavone [31], 5-hydroxy-3,3',4',6,7-pentamethoxyflavone [32], artemitin, rutin, glycosides of quercetin [33], chlorogenic, caffeic acids [33]; bitter principles artamarin, artamaridin, artamaridin, artamarinin [34], Quebrachitol [35]; 24-zeta-Ethylcholesta-7,22-Dien-3-β-ol [36].
<i>A. asiatica</i>	Essential oil containing terpene and a terpene alcohol [37], 1,8-cineole, selin-11-en-4-a-ol, monoterpene alcohols [38], camphor, borneol, bornyl acetate [5,39]; flavone eupatilin (5,7-dihydroxy-3',4', 6-trimethoxyflavone) [40,41]; alkaloids [42]; artemisolide [17].

The ethanol extract of *A. asiatica* has been reported to inhibit inflammatory activation of mouse microglial cells as determined by the production of nitric oxide and the expression of inducible nitric oxide synthase and inflammatory cytokine. The extract also protected nerve growth factor-differentiated PC12 cells against microglial cytotoxicity^[19]. The essential oil 1,8-cineole and selin-11-en-4a-ol, and monoterpene alcohol fraction isolated from *A. asiatica* exhibited antibacterial and antifungal activity against *Bacillus subtilis*, *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Candida albicans*, *Rhodotorula rubra* and *Aspergillus fumigatus*. The monoterpene alcohol fraction showed the highest antibacterial activity^[38]. 4,5-dihydroxy-3',6,7-trimethoxyflavone isolated from *A. asiatica* has been reported to reduce micromolecular amyloid- β protein-induced oxidative cell stress in PC12 cells^[57].

DISCUSSION AND CONCLUSION

Since the beginning of the human civilization, man utilizes native and exotic plants for healing purposes, whose knowledge is transmitted by oral tradition through the generations. Despite the arrival of the industry, in many countries, the majority of the population does not have access to medicines, thus making use of home preparations to cure various health problems. The considerations made on the health of all the people of the world during the declaration of Alma-Ata in 1978, was the milestone that made the World Health Organization (WHO) stress the need, to develop the medicinal plants in the world in the therapeutic scope. Presently, there are different programs supported by WHO, which integrate several preventive and therapeutic aspects of the diseases.

The present review emphasizes the phytochemical, ethnopharmacological, pharmacological reports and toxicological information on *A. absinthium* and *A. asiatica*. Terpenoids, flavonoids, coumarins, caffeoylquinic acids and sterols constitute major classes of phytoconstituents of the genus *Artemisia*. Both *A. absinthium* and *A. asiatica* are rich in terpenoids. *A. absinthium* has been reported to have a broad spectrum of inhibitory activity against a variety of microorganisms due to presence of essential oil^[22]. 5,6,3',5'-tetramethoxy 7,4'-hydroxyflavone, a flavone isolated from *A. absinthium* has been reported to exhibit *in vitro* anti-inflammatory activity^[31]. *A. absinthium* has been studied for cognitive enhancement because of its nicotinic and muscarinic receptor activity in homogenates of human cerebral cortical membranes. Methanol extract of *A. absinthium* enhanced neurite outgrowth induced by nerve growth factor and PC12D cells^[46]. In addition, free-radical scavenging activity of *A. absinthium* extracts has been reported^[47].

DA-9601, a standardized extract of *A. asiatica* exhibits hepatoprotective and chemopreventive effect^[53]. The essential oil 1,8-cineole and selin-11-en-4a-ol, and monoterpene alcohol fraction isolated from *A. asiatica* exhibited antibacterial and antifungal activity^[54]. The ethanol extract of *A. asiatica* has been reported to exhibit anti-inflammatory activity. Eupatilin a pharmacologically active ingredient isolated from *A. asiatica*, has been evaluated for apoptosis-inducing capability in cultured human HL-60 cells^[16].

Despite a long tradition of use of *A. absinthium* and *A. asiatica* for treatment of various ailments, little pharmacological work has been carried out to prove their traditional claims especially in central nervous system (CNS) disorders. Keeping in view the traditional, sporadic phytochemical and pharmacological reports, low toxicity, these species seem to hold great potential for in depth investigation for various biological activities, especially their effects on CNS.

REFERENCES

1. Sanjay J, Satyaendra S, Satish, Sumbhate S. Recent trends in *Curcuma Longa* Linn. Phcog Mag, 1, 2007, 119-128.
2. Bhagwati U. Utilization of medicinal plants by the rural women of Kulu, Himachal Pradesh. Indian J Trad Knowledge, 2, 2003, 366-370.
3. Dahanukar SA, Kulkarni AR, Rege NN. Pharmacology of medicinal plants and natural products. Indian J Pharmacol, 32, 2000, S81-S118.
4. Watson LE, Bates PL, Evans TM, Unwin MM, Estes JR. Molecular phylogeny of Subtribe Artemisiinae (Asteraceae), including *Artemisia* and its allied and segregate genera. BMC Evol Biol, 2, 2002, 17.
5. Kordali S, Cakir A, Mavi A, Kilic H, Yildirim A. Screening of chemical composition and antifungal and antioxidant activities of the essential oils from three Turkish *Artemisia* species. J Agric Food Chem, 53, 2005, 1408-1416.
6. Ribnicky DM, Poulev A, Watford M, Cefalu WT, Raskin I. Antihyperglycemic activity of TarralinTM, an ethanolic extract of *Artemisia dracunculus* L. Phytomedicine, 13, 2006, 550-557.
7. Mehrdad I, Seyed AE, Meysam MS. Detection of sesquiterpene lactones in ten *Artemisia* species population of Khorasan provinces. Iranian J Basic Medi Sci, 10, 2007, 183-188.
8. Koul MK. Medicinal plants of Kashmir and Ladakh, temperate and cold arid Himalaya Indus Publishing Company, FS-5, Tagore Garden, New Delhi, 1997, 102.
9. Wake G, Court J, Pickering A, Lewis R, Wilkins R, Perry E. CNS acetylcholine receptor activity in European medicinal plants traditionally used to improve failing memory. J Ethnopharmacol, 69, 2000, 105-114.
10. Howes MR, Perry NS, Houghton PJ. Plants with traditional uses and activities, relevant to the management of Alzheimer's disease and other cognitive disorders. Phytother Res, 17, 2003, 1-18.
11. Guarrera PM. Traditional phytotherapy in central Italy (Marche, Abruzzo, and Latium). Fitoterapia, 76, 2005, 1-25.
12. Gilani AH, Janbaz K. Preventive and curative effects of *Artemisia absinthium* on acetaminophen and CCl4-induced hepatotoxicity. General Pharmacol, 26, 1995, 309-315.
13. Muto T, Watanabe T, Okamura M, Moto M, Kashida Y, Mitsumori K. Thirteen-week repeated dose toxicity study of wormwood (*Artemisia absinthium*) extract in rats. J Toxicol Sci, 28, 2003, 471-478.
14. Zhang W, Luo S, Fang F. Total synthesis of absinthin. J Am Chem Soc, 127, 2005, 18-19.
15. Harendra SP, Gang L, Ming QW. A new dawn for the use of traditional Chinese medicine in cancer therapy. Mole Cancer, 8, 2009, 21.
16. Seo HJ, Surh YJ. Eupatilin, a pharmacologically active flavone derived from *Artemisia* plants, induces apoptosis in human promyelocytic leukemia cells. Mutation Res, 496, 2001, 191-198.
17. Reddy AM, Lee JY, Seo JH, Kim BH, Chung EY, Ryu SY, Kim YS, Lee CK, Min KR, Kim Y. Artemisolide from *Artemisia asiatica*: nuclear factor-kappa-B (NF-kappaB) inhibitor suppressing prostaglandin E₂ and nitric oxide production in macrophages. Arch Pharmacol Res, 29, 2006, 591-597.
18. Kim HS, Kundu JK, Lee JS, Oh TY, Na HK, Surh YJ. Chemopreventive effects of the standardized extract (DA-9601) of *Artemisia asiatica* on azoxymethane-initiated and dextran sulfate sodium-promoted mouse colon carcinogenesis. Nutr Cancer, 60, 2008, 90-97.
19. Lim BO, Chung HG, Lee WH, Lee HW, Suk K. Inhibition of microglial neurotoxicity by ethanol extract of *Artemisia asiatica* Nakai. Phytother Res, 22, 2008, 279-282.

20. Arnold WN. Absinthe. *Sci Am*, 260, 1989, 112-117.
21. Daise LL, Daniela SA, Celuta SA, Paul PK. Screening of chemical composition, antimicrobial and antioxidant activities of *Artemisia* essential oils. *Phytochemistry*, 69, 2008, 1732-1738.
22. Juteau F, Jerkovic I, Masotti V, Milos M, Mastelic J, Bessiere JM, Viano J. Composition and antimicrobial activity of the essential oil of *Artemisia absinthium* from Croatia and France. *Planta Med*, 69, 2003, 158-161.
23. Goryaev MI, Bazalitskaya VS, Lishtvanova LN. The terpene portion of the essential oil from *Artemisia absinthium*. *Zhurnal Prikladnoi Khimii*, 35, 1962, 2799-2802.
24. Herout V, Sorm F. Constitution of prochamazulenogen, natural precursor of chamazulene in *Artemisia absinthium*. *Collection of Czechoslovak Chemical Communications*, 19, 1954, 792-797.
25. Rezaeinodehi A, Khangholi S. Chemical composition of the essential oil of *Artemisia absinthium* growing wild in Iran. *Pakistan J Biolog Sci*, 11, 2008, 946-949.
26. Anne O, Ain R, Elmar A, Mati M, Tiiu K. Composition of the essential oil of *Artemisia absinthium* L. of different geographical origin. *Proc Estonian Acad Sci Chem*, 55, 2006, 155-165.
27. Leung AY. *Encyclopedia of Common Natural Ingredients Used in Food, Drugs, and Cosmetics*. New York, NY, J Wiley and Sons, 1980.
28. Slepetyts J. Biology and biochemistry of wormwood. Accumulation dynamics of tannins, ascorbic acid, and carotene. *Liet TSR Mokslu Akad Darb Ser C*, 1, 1975, 43-48.
29. Greger H, Hofer O. New unsymmetrical substituted tetrahydrofuran lignans from *Artemisia absinthium*. Assignment of the relative stereochemistry by lanthanide induced chemical shift. *Tetrahedron*, 36, 1980, 3551-3558.
30. Gambelunghe C, Melai P. Absinthe: enjoying a new popularity among young people? *Forensic Sci Int*, 130, 2002, 183-186.
31. Lee HG, Kim H, Oh WK. Tetramethoxy hydroxyflavone p-7F downregulates inflammatory mediators via the inhibition of nuclear factor kappa B. *Ann N Y Acad Sci*, 1030, 2004, 555-568.
32. Cekan Z, Herout V. Plant substances. IV. Isolation of 5-hydroxy-3,3',4',6,7-pentamethoxyflavone from *Artemisia absinthium*. *Collection of Czechoslovak Chemical Communications*, 21, 1956, 79-83.
33. Oswiecimska M, Polak A, Seidl O, Sendra J. Comparative study of chromatograms of the flavonoid fractions from herbs of some species of the genus *Artemisia*. *Dissertationes Pharmaceuticae*, 17, 1965, 503-511.
34. Schenck GS, Nuscha E. The bitter principle of *Artemisia absinthium*. *Archiv der Pharmazie und Berichte der Deutschen Pharmazeutischen Gesellschaft*, 289, 1956, 1-8.
35. Plouvier V. The presence of quebrachitol in some *Artemisia* species. *Annales Pharmaceutiques Francaises*, 7, 1949, 192-195.
36. Ikram M, Shafi N, Mir I, Do MN, Nguyen P, Le Quesne PW. 24zeta-Ethylcholesta-7,22-Dien-3beta-ol: A Possibly Antipyretic Constituent of *Artemisia absinthium*. *Planta Med*, 53, 1987, 389.
37. Kaku T, Yosimura, K. *Artemisia asiatica* Nakai. I. Chemical studies. *Pharmacology*, 11, 1938, 115.
38. Kalembe D, Kusewicz D, Swiader K. Antimicrobial properties of the essential oil of *Artemisia asiatica* Nakai. *Phytothe Res*, 16, 2002, 288-291.
39. Perez-Alonso MJ, Velasco-Negueruela A, Pala-Paul J, Sanz, J. Variations in the essential oil composition of *Artemisia pedemontana* gathered in Spain: chemotype camphor-1,8-cineole and chemotype davanone. *Biochem Syst Ecol*, 31, 2003, 77-84.
40. Kim DH, Na-Hye K, Oh TY, Kim WB, Surh YJ. Eupatilin, a pharmacologically active flavone derived from *Artemisia* plants, induces cell cycle arrest in ras-transformed human mammary epithelial cells. *Biochem Pharmacol*, 68, 2004, 1081-1087.
41. Kim MJ, Kim DH, Na-Hye K, Oh TY, Shin CY, Surh YJ. Eupatilin, a pharmacologically active flavone derived from *Artemisia* plants, induces apoptosis in human gastric cancer (AGS) cells. *J Environmental Pathol, Toxicol Oncol*, 24, 2005, 261-269.
42. Heo HJ, Yang HC, Cho HY, Hong B, Lim ST, Park HJ, Kim KH, Kim HK, Shin DH. Inhibitory effect of *Artemisia asiatica* alkaloids on acetylcholinesterase activity from rat PC12 cells. *Mole Cells*, 10, 2000, 253-262.
43. Khattak SG, Gilani SN, Ikram M. Antipyretic studies on some indigenous Pakistani medicinal plants. *J Ethnopharmacol*, 14, 1985, 45-51.
44. Wake G, Court J, Pickering A, Lewis R, Wilkins R, Perry E. CNS acetylcholine receptor activity in European medicinal plants traditionally used to improve failing memory. *J Ethnopharmacol*, 69, 2000, 105-114.
45. Meschler JP, Howlett AC. Thujone exhibits low affinity for cannabinoid receptors but fails to evoke cannabimimetic responses. *Pharmacol Biochem Behav*, 62, 1999, 473-480.
46. Li Y, Ohizumi Y. Search for constituents with neurotrophic factor-potentiating activity from the medicinal plants of Paraguay and Thailand. *Yakugaku Zasshi*, 124, 2004, 417-424.
47. Jasna M, Canadanovic B, Sonja MD, Gordana SC, Vesna TT. Free-radical scavenging activity of wormwood (*Artemisia absinthium* L) extracts. *J Sci Food Agric*, 85, 2004, 265-272.
48. Tariq KA, Chishti MZ, Ahmad F, Shawl AS. Anthelmintic activity of extracts of *Artemisia absinthium* against ovine nematodes. *Veterinary Parasitol*, 160, 2009, 83-88.
49. Valdes AFC, Martinez JM, Lizama RS, VM, Cos PML. *In vitro* anti-microbial activity of the Cuban medicinal plants Simarouba glauca DC, *Melaleuca leucadendron* L and *Artemisia absinthium* L. *Memorias do Instituto Oswaldo Cruz*, 103, 2008, 615-618.
50. Caner A, Doskaya M, Degirmenci A, Can H, Baykan S, Uner A, Basdemir G, Zeybek U, Guruz Y. Comparison of the effects of *Artemisia vulgaris* and *Artemisia absinthium* growing in western Anatolia against trichinellosis (*Trichinella spiralis*) in rats. *Exp Parasitol*, 1, 2008, 173-179.
51. Chemesova II, Belenovskaya LM, Stukov AN. Antitumor activity of flavonoids from some species of *Artemisia* L. *Rastitel'nye Resursy*, 23, 1987, 100-103.
52. Donald DV. Absinthium: a nineteenth-century drug of abuse. *J Ethnopharmacol*, 1981, 4, 337-343.
53. Ryu BK, Ahn BO, Oh TY, Kim SH, Kim WB, Lee EB. Studies on protective effect of DA-9601, *Artemisia asiatica* extract, on acetaminophen- and CCl4-induced liver damage in rats. *Arch Pharmacol Res*, 21, 1998, 508-513.
54. Kim JY, Kim DY, Lee YS, Lee BK, Lee KH, Ro JY. DA-9601, *Artemisia asiatica* herbal extract, ameliorates airway inflammation of allergic asthma in mice. *Mol Cells*, 22, 2006, 104-112.
55. Oh TY, Ahn GJ, Choi SM, Ahn BO, Kim WB. Increased susceptibility of ethanol-treated gastric mucosa to naproxen and its inhibition by DA-9601, an *Artemisia asiatica* extract. *World J Gastroenterol*, 11, 2005, 7450-7456.
56. Park SW, Oh TY, Kim YS, Sim H, Park SJ, Jang EJ, Park JS, Baik HW, Hahm KB. *Artemisia asiatica* extracts protect against ethanol-induced injury in gastric mucosa of rats. *J Gastroenterol Hepatol*, 23, 2008, 976-984.
57. Heo HJ, Cho HY, Hong B, Kim HK, Kim EK, Kim BG, Shin DH. Protective effect of 4',5-dihydroxy-3',6,7-trimethoxyflavone from *Artemisia asiatica* against Abeta-induced oxidative stress in PC12 cells. *Int J Exp Clin Investigation*, 8, 2001, 194-201.

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